Light-inhibition of dark respiration in Lemna minor L.

Autor(en): **Fuhrer, Jürg**

Objekttyp: **Article**

Zeitschrift: **Botanica Helvetica**

Band (Jahr): **93 (1983)**

Heft 1

PDF erstellt am: **24.09.2024**

Persistenter Link: https://doi.org/10.5169/seals-65235

Botanica Helvetica *93:* 67-75 (1983)

Light-Inhibition of Dark Respiration in *Lemna minor* L.[1)]

Jürg Fuhrer

Pflanzenphysiologisches Institut, Universität Bern, Switzerland

Received January 1983

Abstract

Fuhrer, J. 1983. Light-inhibition of dark respiration in *Lemna minor* L. Bot. Helv. *93:* 67-75. Based on the oxygen-dependence of mesophyll resistance and carbon dioxide compensation concentration, the fraction of dark respiration not inhibited by light was derived for *Lemna minor* L. It was observed that approximately 77% of dark respiration was inhibited at light intensities saturating for net photosynthesis at low carbon dioxide concentration. The corresponding rates of «day» respiration were 13.8 and 12.4 μg CO_2 m^{-2} sec^{-1} at 21 and 1% oxygen, respectively. The rate at 1% oxygen (under non-photorespiratory conditions) was in good agreement with the rate of CO_2-release into CO_2-free air. Rates of «day» respiration were nearly constant at light intensities above 100 μE m^{-2} sec^{-1}, but increased rapidly as the light intensity decreased to lower values.

Introduction

Much of the work carried out in this laboratory has dealt with the regulation of carbon assimilation in C_3 plants. Most recently it was oberved that beans growing on ammonium as N-source show faster ^{14}C-labelling in intermediates of the photosynthetic carbon oxidation (PCO) cycle than those growing on nitrate (Marques et al. 1983. This stimulation of the C-flux to glycollate by ammonium most likely also occurs in other plants, such as *L. minor,* where an enhanced activity of glycollate oxidase was found under the same conditions (Emes and Erismann 1982). Currently, an attempt is being made to predict quantitatively carbon flux rates between pools of major intermediates in PCO and PCR (photosynthetic carbon reduction) cycles based on the measured rate of net C-assimilation and the CO_2 compensation concentration (Fuhrer and Erismann, in preparation). These calculations are based on a model of PCO and PCR cycles which integrates the current knowledge of the biochemistry involved (Farquhar et al., 1980). It is the main idea of this model to express C-fluxes on the basis of the amount of C fixed by ribulose-bisphosphate (RuBP) carboxylase, F_{IN}, and the properties of RuBP carboxylase/oxygenase (RUBISCO).

[1)] This work is dedicated to Prof. Dr. K.H. Erismann, in honor of his 60th birthday.

The reaction of RuBP with one mol of O_2 leads to the release of 0.5 mol of CO_2 through glycine decarboxylation. Therefore, net photosynthesis, F_{Net}, is given by

$$F_{Net} = F_{IN} - 0.5\alpha F_{IN} - R_D \qquad (1a)$$

where α is the ratio between oxygenation and carboxylation of RuBP and R_D denotes CO_2 evolution in the light («day» respiration) due to procresses other than glycine decarboxylation. Dark respiratory processes are known to occur in the light, although at a lower rate than in the dark (Mangat et al. 1974).

Equation (la) can be rearranged to calculate F_{IN} from F_{Net}, R_D and α.

$$F_{IN} = \frac{F_{Net} + R_D}{1 - 0.5\,\alpha} \qquad (1b)$$

To be able to use accurate rates of F_{IN} as the model's input variable, values for R_D have to be known. In this paper, the fraction of dark respiration not inhibited by light is calculated from the O_2-dependence of both, the mesophyll resistance and the CO_2 compensation concentration. Rates were determined at different light intensities to examine whether R_D forms an increasing portion of C-fluxes with decreasing light intensity, as predicted by Farquhar's model.

Theoretical considerations for the calculation of «day» respiration rates

The calculation of R_D was based on a simple model of CO_2 exchange presented by Peisker and Apel (1980). According to this model, R_D can be calculated from the O_2-dependence of (i) the mesophyll resistance, r_m, and (ii) the CO_2 compensation concentration, τ.

The relationship between r_m and the O_2 concentration can be written as,

$$\beta = \frac{\triangle r_m}{\triangle [O_2]} \qquad (2)$$

and was calculated from values of r_m obtained at 1% and 21%O_2. The mesophyll resistance at each O_2 concentration was derived as the slope of the relationship between F_{Net} and the intercellular CO_2 concentration.

$$r_m = \frac{[CO_2] - \tau - r_s}{F_{Net}} \qquad (3)$$

Resistance to CO_2 exchange, r_s, (commonly called «stomatal resistance», but gas exchange in *Lemna minor* is likely to occur through the cuticle as well) was taken as a constant value of 1.6 cm/sec (unpublished observation). Intercellular CO_2 concentrations were calculated according to von Caemmerer and Farquhar (1981).

The CO_2 compensation concentration, τ, depends on the O_2 concentration in a linear way (Forrester et al. 1966),

$$\gamma' = \frac{\triangle \tau}{\triangle [O_2]} \qquad (4)$$

and the whole O_2 dependence of γ' is given by

$$\gamma' = \gamma + \mu \cdot \beta \cdot R_N$$

where R_N is the rate of dark respiration.

This relationship is linear when β, μ and R_N are constant, and γ, the part of the O_2 -dependence which only depends on the kinetic properties of RUBISCO, can be derived as intersection with the ordinate axis (Peisker et al. 1981). Under the different light conditions used in the present study, however, γ, the fraction of R_N not inhibited by light, becomes variable. Therefore, γ had to be derived by extrapolation of the non-linear relationship (see Fig. 2).

For each treatment, $\beta \cdot R_N$ and γ' were determined and used together with the constant value for γ in Eq. (5b) to calculate μ:

$$\mu = \frac{\gamma' - \gamma}{\beta \cdot R_N} \qquad (5b)$$

γ, which did not depend on R_D, was also used to determine the CO_2 compensation concentration in the absence of R_D, τ^*, using Eq. (6) according to Farquhar et al. (1980).

$$\tau^* = [O_2] \cdot \gamma \qquad (6)$$

Materials and Methods

Lemna minor L. were cultivated in aerated Fernbach flasks at 20 °C and ambient CO_2 for seven days on a modified Hutner medium containing 7 mM NO_3^- as nitrogen source (Emes and Erismann 1982). Light intensity was 250 μE m^{-2} sec^{-1} provided by fluorescence bulbs placed beneath and above the culture flasks.

Rates of net photosynthesis (F_{Net}) and dark respiration (R_N) were determined after transfer of the culture to a round, stainless steel cuvette (with a glas window build into the removable cover) which was part of an open fumigation system (designed by Prof. K.H. Erismann). Air flow through the cuvette was 1 l min^{-1}. Carbon dioxide from a pressure tank was added to CO_2-free and dried ambient air through permeation tubes placed in the gas stream. Individual CO_2 concentrations were obtained by adjusting the length of the permeation tube. Air entering or leaving the cuvette was analyzed for CO_2 with a Siemens IRGA (Ultramat 32) after passing through a condenser at 4°C. Oxygen concentrations other than that of ambient air (21%) were obtained by using commercially purchased O_2/N_2 mixtures (Carba Gas, Liebefeld-Bern) instead of ambient air.

Quantum yield of net photosynthesis under different CO_2 and O_2 conditions was determined according to Ehleringer and Björkmann (1977). Total leaf area of a culture was estimated on a photographic picture with a planimeter.

The open IRGA system was adapted to measure the CO_2 compensation concentration, τ, at 25 °C. The gas from the sample cell of the IRGA was recycled through the plant cuvette back to the IRGA by a diaphragm pump to create a closed system. Before closure, the system was flushed with the appropriate O_2/N_2 mixture and 578 ng cm^{-3} CO_2.

Light (1200 μE m^{-2} sec^{-1} PHAR, 400-750 nm) provided by a 1000 W quartz iodide lamp (Philips 12013 R) placed above a tank containing circulating tap water. The cuvette was covered with a plastic bowl containing a 2 cm deep layer of a $CuSO_4$ solution (10 g/500 ml) to reduce heat transfer into the cuvette. Various light intensities were obtained by placing metal screens with different size holes above the cuvette. Light intensities were measured with a Quanta spectrometer (Techtum QSM 2500). The temperature in the cuvette was adjusted to 25 °C by circulating water from a temperature-controlled water bath through an inner compartment in the stainless steel body.

Carbon dioxide evolution in the dark (R_N) was measured after a 20 min light period followed by a 20 min dark period.

Results and Discussion

Net CO_2 uptake rate of *L. minor*, F_{Net}, depended on the light intensity (I) and the CO_2 and O_2 concentrations as shown in Fig. 1A. Table 1 contains the quantum yields determined from the data presented in Fig. 1A. They were in agreement with those from other C_3 plants (Ehleringer and Björkmann 1977).

Table 1: Quantum yields for *Lemna minor* L. at different CO_2 and O_2 concentrations.

	CO_2 concentration (ng cm^{-3})	
	150	478
O_2 concentration (%): 21	0.032	0.068
1	0.080	0.080

A rapid decrease of F_{Net} occurred as I decreased from about 100 μE m^{-2} sec^{-1} to the light compensation point. On the other hand, an increase of τ over the same range of I was observed (Fig. 1B). At I saturating for F_{Net} at low CO_2 concentration, τ was approximately 90 ng cm^{-3} at 21% O_2 and 19 ng cm^{-3} at 1% O_2. The value of τ at 21% O_2 was slightly higher than of wheat leaves (Feller and Erismann 1978, Peisker and Apel 1975), but lower than that of adult bean leaves (Peisker et al. 1981). A similar increase of τ below 260 μE m^{-2} sec^{-1} (equivalent to 84 μE m^{-2} sec^{-1} when using an IR-filter as in this study) was reported for bean leaves of different age (Catský and Tichá 1979). The observed increase of τ under condition of inhibited photorespiration (1% O_2) indicated the apparent effect of «day» respiration, R_D. According to Farquhar et al. (1980), τ depends on the properties of RUBISCO, the O_2 concentration and the rate of R_D. A linear correlation between τ and $R_D / V_{c,max}$, with $V_{c,max}$ as maximal carboxylation velocity, can be observed (Peisker et al. 1981).

Table 2: Mean values for r_m at 21% and 1% O_2, respectively, the O_2-dependence of the carboxylation resistance, β, the product of β and the rate of dark respiration, R_N, the O_2-dependence of τ, γ', and the difference between γ' and the O_2-dependence of the CO_2 compensation concentration in the absence of «day» respiration, γ, in *Lemna minor* L. at different light intensities. Standard errors were generally within ± 10% of the mean value (not shown).

I (μE m^{-2} sec^{-1})	r_m (21%) (sec m^{-1})	r_m (1%) (sec m^{-1})	β (sec m^{-2} kg^{-1})	$\beta \cdot R_N$ (g kg^{-1})	γ' (g kg^{-1})	$\gamma' - \gamma$ (g kg^{-1})
380	1313	1255	215	0.0129	0.270	0.003
150	1710	1571	515	0.0309	0.276	0.009
90	2461	2238	826	0.0496	0.290	0.023
60	4000	3724	1022	0.0613	0.301	0.038
50	5299	4892	1248	0.0749	0.316	0.049

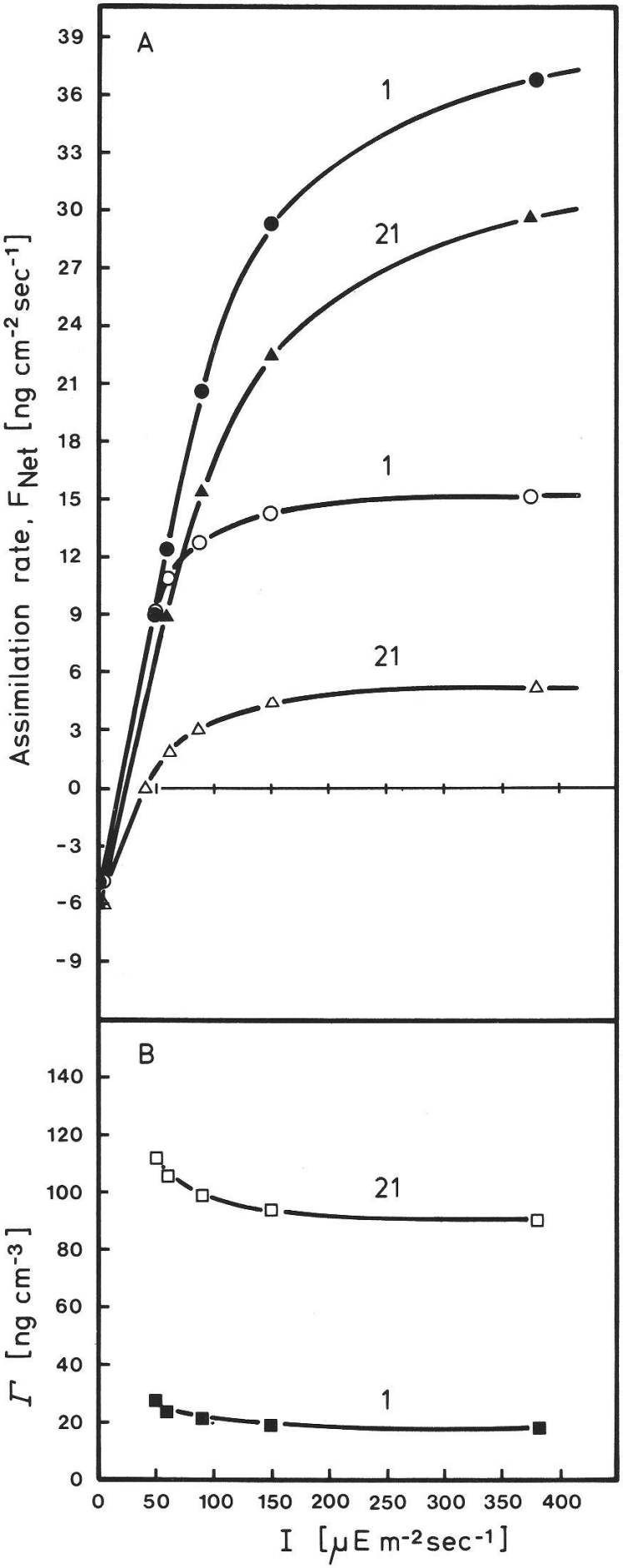

Fig.1: A. Rate of net CO_2 assimilation, F_{Net}, versus light intensity, I, in *L. minor* L. measured at 528 ng cm^{-3} (closed symbols) or 150 ng cm^{-3} (open symbols) external CO_2 concentration, and at either 21% or 1% O_2. Negative values of F_{Net} indicate CO_2 evolution.
B. Carbon dioxide compensation, concentration, τ, versus light intensity, I, in *L. minor* L. at 21% or 1% O_2.

The inital slope of the relationship between F_{Net} and the intercellular CO_2 concentration was used as a measure for the mesophyll resistance to CO_2, r_m (Table 2). Values obtained with air containing 1% and 21% O_2 were used to derive the slope of the dependence of r_m on O_2, β. Values for β largely depended on I (Table 2).

The rate of dark respiration, R_N, measured after a 20 min dark period following 20 min of light, was 60 ± 10 μg CO_2 m^{-2} sec^{-1} at 21% O_2, similar to the rate determined for wheat leaves under comparable experimental conditions (Peisker and Apel 1975), and 50 ± 6 μg CO_2 m^{-2} sec^{-1} at 1% O_2. The calculated factor $\beta \cdot R_N$ is listed in Table 2 for the different light intensities used. This factor was lowest at light intensities saturating for F_{Net} at low CO_2 concentrations (see Fig. 1A).

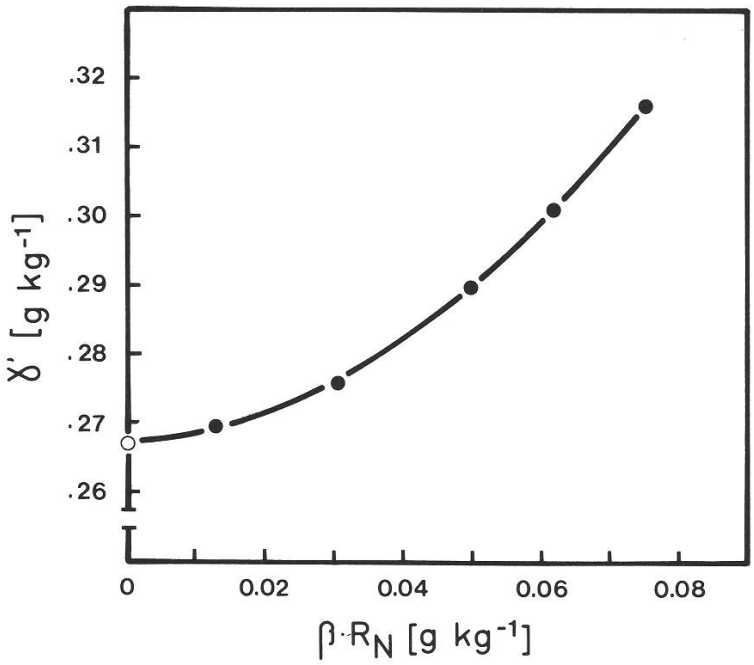

Fig. 2: Relationship between the O_2-dependence of the CO_2 compensation concentration, γ', and the product of the O_2-dependence of the carboxylation resistance, β, and the CO_2 evolution in darkness, R_N, in *L. minor* L. The intersection with the ordinate axis, (open circle), γ, was found by extrapolation.

Values for $\beta \cdot R_N$ were plotted against those found for γ' at the diffrent light intensities Fig. 2). The non-linear regression line was extrapolated to $\beta \cdot R_N = O$. At this point, γ' equaled γ (0.276 g kg^{-1}). γ should not differ greatly between C_3 species and should not be influenced by varying I. In fact, Peisker and Apel (1980) reported 0.26 g kg^{-1} for wheat leaves at 23 °C and Peisker et al. (1981) found 0.308 g kg^{-1} for primary leaves of beans at 28 °C. Charles-Edwards (1978) listed a series of values around 0.24 g kg^{-1} for a variety of C_3 plants at 25 °C. The accuracy of the value for *L. minor* reported here is therefore satisfactory, considering the possible limitations for the model of Peisker and Apel (1980) at high CO_2 concentrations and the incertainity in deriving γ by extrapolation.

The value for γ was used in Eq. (6) to calculated τ^*, the CO_2 compensation concentration in the absence of «day» respiration. At 21% O_2, τ^* was 71.8 ng cm^{-3}. This concentration only depends upon the characteristics of RUBISCO (Farquhar et al. 1980). A similar value for different C_3 species could be expected. Farquhar et al. (1980) calculated 55 ng cm^{-3} based on *in vitro* data from the spinach chloroplast enzyme, and data from Badger and Andrews (1974) suggest 81 ng cm^{-3}. The difference meight be due to differences in the *in vitro* assays used by these authors.

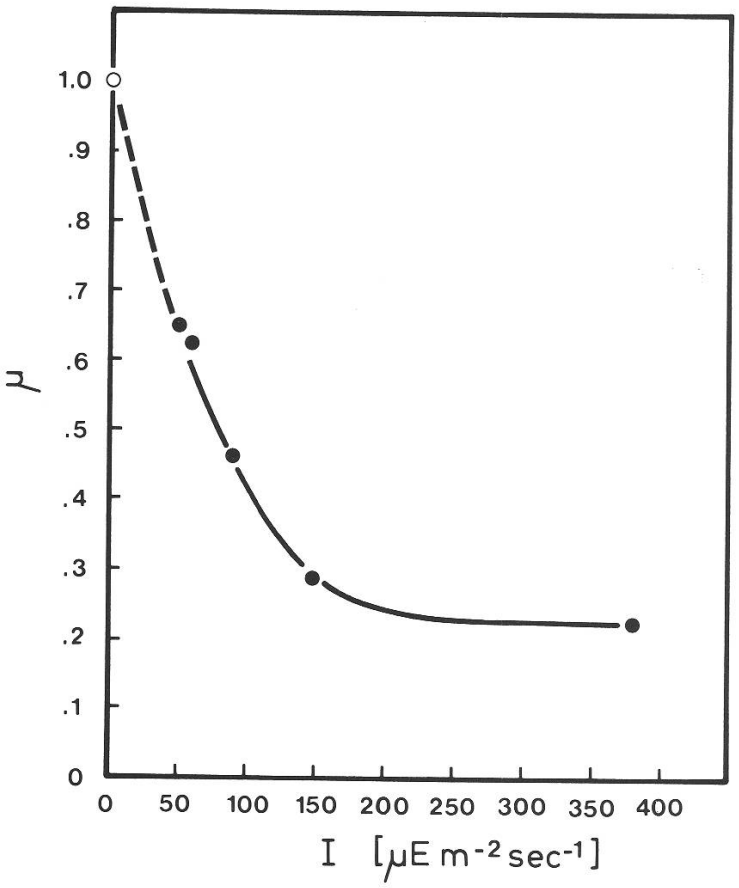

Fig. 3: The fraction of dark respiration not inhibited by light, γ, versus light intensity, I, in *L. minor* L.

Based on the values for $\beta \cdot R_N$ and γ'-γ given in Table 2, the fraction of R_N not inhibited by light, μ, was calculated according to Eq. (5b). Values obtained at each light intensity are plotted in Fig. 3. It was found that μ was about 0.23 at high light intensity, which means that 77% of R_N was inhibited by light, leaving a rate of R_D of 13.8 μg CO_2 m^{-2} sec^{-1}. This equaled 7.7% of F_{Net} at ambient CO_2 and O_2 concentrations. Farquhar et al. (1980) assumed a comparable value of 5.8%. A smaller light inhibition of R_N (66%) was observed in wheat leaves at 500 μE m^{-2} sec^{-1} (Peisker and Apel 1980). Values for bean leaves fluctuated between 75% and 20% during ontogenesis (Peisker et al. 1981). Mangat et al. (1974) derived 75% inhibition for bean leaves based on ^{14}C labelling experiments.

At I limiting for F_{Net}, μ increased. A value of 39.2 μg CO_2 m^{-2} sec^{-1} for R_D was calculated from μ (= 0.654) at 50 μE m^{-2} sec^{-1}. The broken line in Fig. 3 indicates a possible extrapolation below the light compensation point to complete darkness where R_D equals R_N. The increase of μ with decreasing I agrees with the prediction by the model of Farquhar et al. (1980). This observation underlines the possibility that «day» respiration depends on ATP supplied by photophosphorylation which would be low at light intensities limiting for photosynthesis (Mangat et al. 1974).

Assuming thatR_D was inhibited by low O_2 concentration to a similar extent as R_N (10%), R_D at 1% O_2 was 12.4 μg CO_2 m^{-2} sec^{-1}. On the other hand, photorespiratory CO_2-release is likely to be very small at 1% O_2. Therefore, the rate of CO_2-release into CO_2-free air at 1% O_2 should be similar to the rate of R_D. In fact, a value of 12.6 ± 3.5 (S.E.) μg CO_2 m^{-2} sec^{-1} was determined at high light intensities. This agreement indicates that R_D can be estimated in CO_2 gas exchange experiments from the CO_2 release into CO_2-free air under non-photorespiratory conditions.

Zusammenfassung:

Auf der Grundlage der Sauerstoffabhängigkeit des Mesophyllwiderstandes und der Kohlendioxid-Kompensationskonzentration wurde der Anteil der Dunkelatmung von *Lemna minor* bestimmt, der nicht durch Licht gehemmt wird. Es wurde beobachtet, daß ungefähr 77% der Dunkelatmung durch Lichtintensitäten, die für die Netto-Photosynthese bei geringer CO_2-Konzentration sättigend sind, gehemmt werden. Die entsprechenden Raten der «Licht»-Atmung betrugen 13,8 und 12,4 $\mu g\ CO_2\ m^{-2}\ sec^{-1}$ bei 21%, bzw. 1% Sauerstoff. Die Rate bei 1% Sauerstoff (bei gehemmter Photorespiration) stimmte gut mit der Rate der CO_2-Abgabe in CO_2-freie Luft überein. Unter Lichtintensitäten über $100 \mu E\ m^{-2}\ sec^{-1}$ waren die Raten der «Licht»-Atmung nahezu konstant, stiegen aber an, wenn die Lichtintensität unter diesen Wert absank.

This work has been supported by Swiss National Science Foundation grant 3.067-0.81. The author wishes to express his appreciation to Prof. K.H. Erismann for his support and constant interest and his help with the gas exchange experiments.

References

Badger M.R. and T.J. Andrews 1974. Effects of CO_2, O_2 and temperature on a high-affinity form of ribulose diphosphate carboxylase-oxygenase from spinach. Biochem. Biophys. Res. Commun. *60:* 204-210.

Caemmerer S. von and G.D. Farquhar 1981. Some relationships between the biochemistry of photosynthesis and the gas exchange of leaves. Planta *153:* 376-387.

Čatský J. and Tichá 1979. CO_2 compensation concentration in bean leaves: Effect of photon flux density and leaf age. Biol. Plant. *21:* 361-364.

Charles-Edwards D.A. 1978. Leaf carbon dioxide compensation points at high light flux densities. Ann. Bot. *42:* 733-739.

Ehleringer J. and O. Björkmann 1977. Quantum yields for CO_2 uptake in C_3 and C_4 plants. Plant Physiol. *59:* 86-90.

Emes M.J. and K.H. Erismann 1982. The influence of the nitrogen supply on the structure and activity of glycolate oxidase in *Lemna minor* L. Plant. Sci. Lett. *27:* 103-109.

Farquhar G.D., S. von Caemmerer and J.A. Berry 1980. A biochemical model of photosynthetic CO_2 assimilation in leaves of C_3 species. Planta *149:* 78-90.

Feller U. and K.H. Erismann 1978. Veränderungen des Gaswechsels und der Aktivitäten proteolytischer Enzyme während der Seneszenz von Weizenblättern *(Triticum aestivum* L.). Z. Pflanzenphysiol. *90:* 235-244.

Forrester M.L., G. Krotkov and D.C. Nelson 1966. Effect of oxygen on photosynthesis, photorespiration and respiration in detachted leaves. I. Soybean. Plant Physiol. *41:* 422-427.

Mangat B.S., W.B. Levin and R.G.S. Bidwell 1974. The extent of dark respiration in illuminated leaves and its control by ATP levels. Can. J. Bot. *52:* 673-681.

Marques I.A., M.J. Oberholzer and K.H. Erismann 1983. Effects of different inorganic nitrogen sources on photosynthetic carbon metabolism in primary leaves of non-nodulated *Phaseolus vulgaris* L. Plant. Physiol. *71:* 555-561.

Peisker M. and P. Apel 1980. Dark respiration and the effect of oxygen on CO_2 compensation concentration in wheat leaves. Z. Pflanzenphysiol. *100:* 389-395.

Peisker M. and P. Apel 1975. Influence of oxygen on photosynthesis and photorespiration in leaves of *Triticum aestivum* L. 1. Relationship between oxygen concentration, CO_2 compensation point, and intracellular resistance to CO_2 uptake. Photosynthetica *9:* 16-23.

Peisker M., I. Tichá and J. Čatský 1981. Ontogenetic changes in the internal limitations to bean-leaf photosynthesis. 7. Interpretation of the linear correlation between CO_2 compensation concentration and CO_2 evolution in darkness. Photosynthetica *15:* 161-168.

Dr. J. Fuhrer
Pflanzenphysiol. Institut der Universität
Altenbergrain 21
CH-3013 Bern (Switzerland)