Zeitschrift:	Archives des sciences et compte rendu des séances de la Société			
Herausgeber:	Société de Physique et d'Histoire Naturelle de Genève			
Band:	52 (1999)			
Heft:	3			
Artikel:	Further 31P-NMR evidence for phosphoramide bonding of nucleobases : by MG2+ enhanced nucleophilic attack on cyclic triphosphate			
Autor:	Turian, Gilbert / Rivara-Minten, Elisabeth / Cattaneo, Arlette			
DOI:	https://doi.org/10.5169/seals-740115			

Nutzungsbedingungen

Die ETH-Bibliothek ist die Anbieterin der digitalisierten Zeitschriften auf E-Periodica. Sie besitzt keine Urheberrechte an den Zeitschriften und ist nicht verantwortlich für deren Inhalte. Die Rechte liegen in der Regel bei den Herausgebern beziehungsweise den externen Rechteinhabern. Das Veröffentlichen von Bildern in Print- und Online-Publikationen sowie auf Social Media-Kanälen oder Webseiten ist nur mit vorheriger Genehmigung der Rechteinhaber erlaubt. <u>Mehr erfahren</u>

Conditions d'utilisation

L'ETH Library est le fournisseur des revues numérisées. Elle ne détient aucun droit d'auteur sur les revues et n'est pas responsable de leur contenu. En règle générale, les droits sont détenus par les éditeurs ou les détenteurs de droits externes. La reproduction d'images dans des publications imprimées ou en ligne ainsi que sur des canaux de médias sociaux ou des sites web n'est autorisée qu'avec l'accord préalable des détenteurs des droits. <u>En savoir plus</u>

Terms of use

The ETH Library is the provider of the digitised journals. It does not own any copyrights to the journals and is not responsible for their content. The rights usually lie with the publishers or the external rights holders. Publishing images in print and online publications, as well as on social media channels or websites, is only permitted with the prior consent of the rights holders. <u>Find out more</u>

Download PDF: 01.08.2025

ETH-Bibliothek Zürich, E-Periodica, https://www.e-periodica.ch

Communication présentée à la séance du 21 octobre 1999

FURTHER ³¹P-NMR EVIDENCE FOR PHOSPHORAMIDE BONDING OF NUCLEOBASES BY MG²⁺ ENHANCED NUCLEOPHILIC ATTACK ON CYCLIC TRIPHOSPHATE

ΒY

Gilbert TURIAN*, Elisabeth RIVARA-MINTEN** & Arlette CATTANEO*

Abstract

Further ³¹P-NMR evidence for phosphoramide bonding of nucleobases by Mg²⁺ enhanced nucleophilic attack on cyclic triphosphate. - Mg²⁺ enhanced nucleophilic attack by nitrogen bases, phosphoramido-bonded on the opening cycle of trimetaphosphate detected by ³¹P-NMR spectral changes, is extended from imidazole to purines and pyrimidines.

Key-words: NMR, Nucleobases, Imidazole, Cyclic and Linear Triphosphates

INTRODUCTION

We have recently interpreted the activation by either imidazolides or nucleobases of the carbodiimide (EDC)-catalyzed condensation of linear triphosphate to the cyclic trimetaphosphate as due to the simultaneous opening of the triphosphate cycle products, thereby displacing the chemical equilibrium in favor of further cyclic condensation (TURIAN *et al.*, 1998).

The opening of the triphosphate cycle was conceivably the result of the well known nucleophilic attack of the >NH containing azole bases on one of the phosphoanhydric bonds of the polyphosphate molecules (RABINOWITZ & HAMPAI, 1985). This proposal has led us to the present attempt to obtain direct evidence of phosphoramidic bonding with the nucleobases, all bearing in their molecule the same reactive >NH group (N1 in pyrimidines, N9 in purines).

Such a possible phosphoramide (N-P) bonding of nucleobases would be most interesting in the perspective that the presumed "prebiotic" synthesis of prenucleic polymers might have first short-circuited that of nucleotidic nucleic acids (TURIAN, 1996-1998).

^{*} Laboratoire de Bioénergétique et Microbiologie, Pavillon des Isotopes, 20, boulevard d'Yvoy, CH-1211 Genève 4.

^{**}Laboratoire de Chimie pharmaceutique, Sciences II, Université de Genève, CH-1211 Genève 4.

MATERIALS AND METHODS

"Prebiotic" chemicals were dissolved in 2 ml San Pellegrino mineral water (SP, Milano, Italy) chosen because of its equilibrated light mineral composition and pH (7.7):

10 (5) mM of trimetaphosphate (Sigma, grade III) was dissolved in this solution naturally containing 2.5 mM of Mg^{2+} or enriched with 5 (10) mM of $MgCl_2.6H_2O$ to benefit of the known increased sensitivity of the phosphorus bonds to the nucleophiles in the presence of cationic metals (CORBRIDGE, 1978). The solutions were incubated in the presence of each nucleobase (Sigma, 5 mM except 0.25 mM for guanine) or imidazole (Fluka, 5 mM) from 11 to 18 days in shaken Pyrex capped miniflasks at 25°C.

The percent trimetaphosphate decyclized to linear triphosphate was determined by ³¹P-NMR at 81 MHz on a AC200F Bruker NMR spectrometer using H_3PO_4 as an external reference. Ratios of trimetaphosphate (singlet signalled at -21.5 ppm, VOGEL, 1984) and linear triphosphate (2 resonance peaks, doublet + triplet signals, CALLIS *et al.*, 1957; VAN WAZER, 1958) were measured by integration and normalized on the external reference (capillary containing H_3PO_4 solution).

The nucleophilic bases have been imidazole (efficient nucleophile, CHUNG *et al.*, 1971; RABINOWITZ & HAMPAI, 1978), and the imidazole-ring containing purines (adenine and guanine) or the pyrimidines (cytosine and uracil) also bearing the nucleophilic >NH group.

RESULTS AND DISCUSSION

In the low magnesium controls, all nitrogen bases assayed have provoked a moderate opening of the trimetaphosphate (Table Ia, Fig. 1). By contrast, supplementary Mg²⁺

Nitrogen bases	Reaction times		Nitrogen bases	Reaction times	
	11 d	18 d		11 d	18 d
a) Controls	0*	0**	b) $+ Mg^{2+}$	9	11
Imidazole	12	14	Imidazole	28	35
Adenine	8	10	Adenine	19	22
Guanine	5	7	Guanine°	11	15
Cytosine	7	11	Cytosine	15	17
Uracil	9	8	Uracil	25	21

TABLE I
Decyclization of trimetaphosphate (% - TriMP)

Comparative efficiencies of TriMP (10 mM) decyclization by 5 mM nitrogen bases (imidazole or nucleobases) incubated in mineral water a) native (low Mg^{2+} , controls) or b) enriched in 5 (10) mM Mg^{2+} .

Results calculated from the lowerings of the ratio "height of the ${}^{31}P$ NMR peak of the TriMP singlet (-21.5 ppm) / height of the H₃PO₄ capillary reference", expressed as % of such lowerings averaged from at least 2 series of experiments.

*-** TriP cycles of controls have to be considered as 98% intact (Sigma, grade III) even though they and all decycling values presented have undergone additional 1-2% of further spontaneous cycloTriP openings during incubations.

alone produced additional cycle opening as evidenced by a significant decrease of the -21.5 ppm (Table Ib). This decyclization was presumably due to the straight hydrolysis of an anhydride bond of triphosphate as also occurring with ATP (MAHLER & CORDES, 1969) and polyphosphates (KORNBERG, 1995; KORNBERG *et al.*, 1999).

However, in our experimental conditions, the labilization of the phosphoanhydride bonding by Mg^{2+} ions led to the 2 peak linear triphosphate only (Fig. 1A,B,C), contrarily to the splitting into pyrophosphate + Pi reported by SHABAROVA (1970).

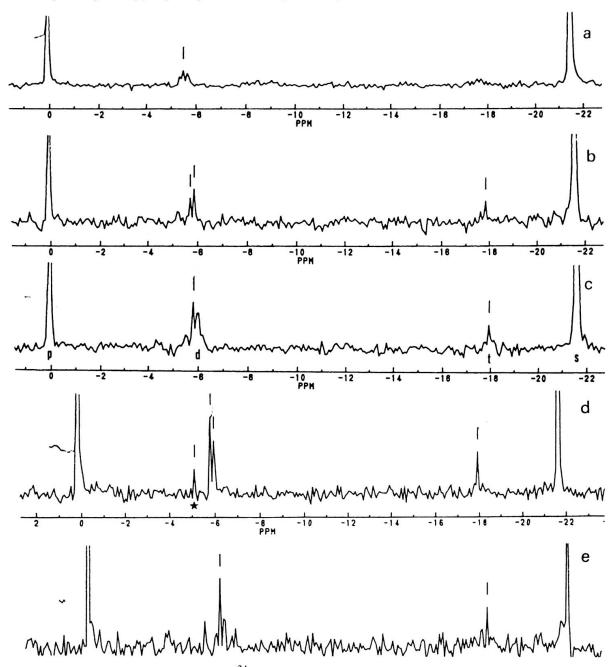


FIG. 1A. Comparative intensities of the ³¹P NMR spectral signals of linear triphosphate (doublet = d + triplet = t) increasingly decyclized (18 days) from 10 mM trimetaphosphate (singlet = s) in mineral water (hydrolytic control a) by nucleophilic attack of the purine **adenine** (5 mM) alone (b) or doped by 5 mM Mg^{2+} ions (d) with hydrolytic control 5 mM Mg^{2+} ions alone (c) and with **imidazole** as optimal reference (e). * = oligophosphate peak ?

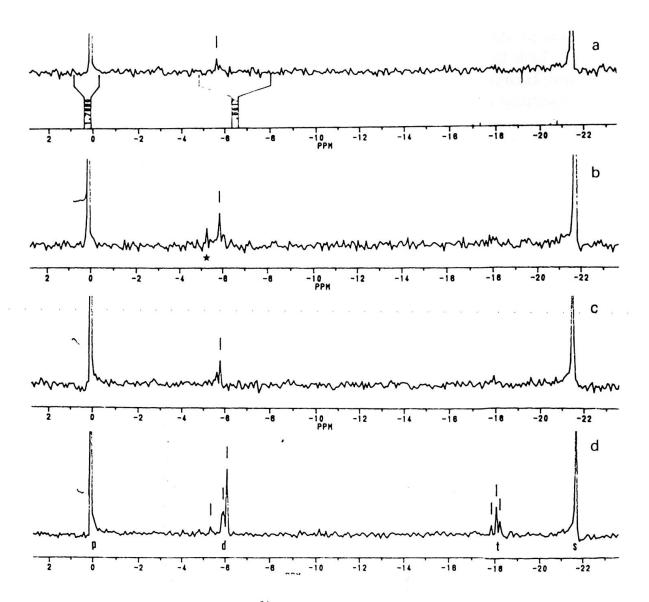


FIG. 1B. Comparative intensities of the ³¹P NMR spectral signals of linear triphosphate (doublet = d + triplet = t) decyclized (18 days) from 10 mM trimetaphosphate (singlet = s) in mineral water (hydrolytic control a) by nucleophilic attack of the pyrimidine **cytosine** (5 mM) alone (b) or doped by 5 mM Mg²⁺ ions (d) with hydrolytic control 5 mM Mg²⁺ ions alone (c).

The most significant decrease of the trimetaphosphate signal was measured in the high Mg2+ solutions incubated with each of the 4 nucleobases and, optimally, with imidazole (Table Ib, Fig. 1A,B). When doped with Mg²⁺ ions, pyrimidines were as effective decycling molecules (Fig. 1B,C) as the imidazolide purines (Fig. 1A). Uracil even showed the sharpest differential when tested with supplementary Mg²⁺ (10 mM).

As for the enhancing effect of Mg^{2+} ions on the cycle opening by all nitrogen bases, it could rather be ascribed to their shielding effect of the charged groups of cyclic phosphate favoring (Westheimer, 1987) - and thereby revealing - their nucleophilic attack by the nitrogen bases (Fig. 2).

Interestingly, a small but sharp peak at \sim -4.5 ppm, visible downstream of the \sim -6 ppm doublets (Fig. 1*) which increased after 18 days incubation, might be identical to

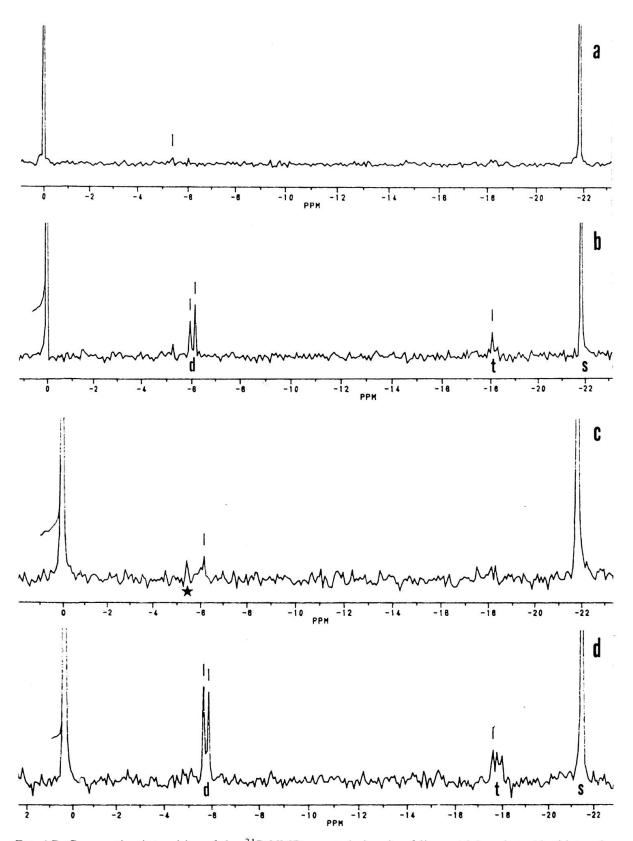


FIG. 1C. Comparative intensities of the ³¹P NMR spectral signals of linear triphosphate (doublet = d + triplet = t) increasingly decyclized (11-18 days) from 5 mM trimetaphosphate (singlet = s) by 10 mM Mg²⁺ ions alone (a,c) and decyclized by nucleophilic attack of the pyrimidine **uracil** (5 mM) doped by 10 mM Mg²⁺ ions (b,d).

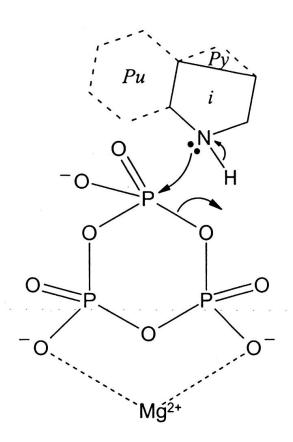


FIG. 2. Nucleophilic attack by the >NH group of nitrogenous bases (i = imidazole; Pu = purines; Py = pyrimidines) on an electrophilic P atom of trimetaphosphate (TriMP), resulting in an open chain or linear triphosphate. Mg²⁺ ions shield the ⁻ charges of the TriMP, thereby favoring its decyclization.

that ascribed by GILLIES *et al.* (1982) to the terminal phosphate of oligo-polyphosphates. Such peak could be considered as the polymerization product -hexo-nono-dodecapolyphosphate- precedingly modellized (Fig. 1 in TURIAN, 1998) as base-bearing riboseless anhydride phosphate polymers which might have been precursors of the phosphoester bonded polyribonucleotides and thereby have assumed the function of some primitive type of replicators during prebiotic evolution.

In conclusion, our experimental results provide a first clue to the elusive problem of the precedence of the "modern" N-C-P, glycosidic-phosphodiester bonds with possible C precursors such as C_2 - C_3 hydroxylated carbon compounds leading to the standard C_5 pyranoses finalized as furanoriboses (ESCHENMOSER, 1999). Such anteriority by direct N-P bondings of nucleobases lined up on polytriphosphates as primal amino acid coding units (TURIAN, 1996) would have been an advantage of simplicity at the onset of pregenetic molecular evolution: it might have provided, by catalytic acylation-phosphorylation entailing peptide bond formation from imidazole triphosphate (RABINOWITZ & HAMPAI, 1985; YAMANAKA *et al.*, 1988) or similarly from our putative nucleobase-triphosphates (adenyl-P, cytidyl-P, etc.), a self-entrained coupling mechanism of primary translation into more of the primally coded peptides.

ACKNOWLEDGEMENTS

We are especially grateful to Professor Jean Tronchet (Pharmaceutical Chemistry Lab.) for availability of his NMR installation (E.R.-M.) and also to Dr. P.-Y. Morgantini (Physical Chemistry Lab., Prof. J. Weber) for the graphics modelling. We also thank Professor Reto Strasser and Dr. Mukti Ojha (Bioenergetics and Microbiology Lab.) for technical facilities (A.C) and Ariane Fehr for competent secretarial assistance.

RÉSUMÉ

NOUVELLE ÉVIDENCE PAR ³¹P-RMN D'UNE LIAISON PHOSPHORAMIDE DES NUCLÉOBASES PAR ATTAQUE NUCLÉOPHILE DU TRIPHOSPHATE CYCLIQUE

L'attaque nucléophile, dopée par les ions Mg²⁺, des bases azotées phosphoramidoliées sur le cycle ouvert du trimétaphosphate a été détectée par les changements de spectres ³¹P-RMN et généralisée de l'imidazole aux purines et pyrimidines.

Mots-clés: RMN, Nucléobases, Imidazole, Triphosphates cycliques et linéaires.

REFERENCES

- CALLIS, C.F., J.R. VAN WAZER, J.R. SHOOLERY & W.A. ANDERSON. 1957. Principles of phosphorus chemistry. III. Structure proofs by nuclear magnetic resonance. J. Amer. Chem. Soc. 79: 2719-2726.
- CHUNG, N.M., R. LOHRMANN, L.E. ORGEL & J. RABINOWITZ. 1971. The mechanism of the trimetaphosphate-induced pyrimidine synthesis. *Tetrahedron* 27: 1205-1210.
- CORBRIDGE, D.E.C. 1978. Phosphorus. An Outline of its Chemistry, Biochemistry and Technology. Elsevier Sci. Publ. Co.
- ESCHENMOSER, A. 1999. Chemical etiology of nucleic acid structure. Science 284: 2118-2124.
- GILLIES, R.J., J.R. ALGER, J.A. DEN HOLLANDER & R.G. SHULMAN. 1982. Intracellular pH measured by NMR: methods and results. *In: Intracellular pH: its Measurements, Regulation and Utilization in Cellular Functions*. Fig. 2, p. 81. Alan R. Liss Inc. New York.
- KORNBERG, A. 1995. Inorganic polyphosphate: toward making a forgotten polymer unforgettable. J. Bacteriol. 177: 491-496.
- KORNBERG, A., N.N. RAO & D. AULT-RICHÉ. 1999. Inorganic polyphosphate: a molecule of many functions. Annu. Rev. Biochem. 68: 89-125.
- MAHLER, H.R. & E.H. CORDES. 1969. Biological Chemistry. Harper Intern. Ed.
- RABINOWITZ, J. & A. HAMPAI. 1978. Influence of imidazole and hydrocyanic acid derivatives on the "possible prebiotic" polyphosphate induced peptide synthesis in aqueous solution. *Helv. Chim. Acta* 61: 1842-1847.
- RABINOWITZ, J. & A. HAMPAI. 1985. Quantitative polyphosphate-induced "prebiotic" peptide formation in H₂O by addition of certain azoles and ions. J. Mol. Evol. 21: 199-201.
- SHABAROVA, Z.A. 1970. Synthetic nucleotide peptides. In: Progress in Nucleic Acid Research and Molecular Biology. DAVIDSON, J.N. & W.G. COHN, Eds. 10: 145-180.
- TURIAN, G. 1996. Polarity at onset of genetic coding. I. Bipolar bondings in the two-step takeover of peptide templates by prenucleic-ribonucleic acids. *Archs Sci. Genève* 49: 213-227.
- TURIAN, G. 1998. Origin of life. I. Recurrent riddles about its genetic coding. Arch. Sci. Genève 51: 311-323.
- TURIAN, G., E. RIVARA-MINTEN & A. CATTANEO. 1998. Similar ³¹P-NMR signals emitted by imidazole and the nucleobases presumably phosphoramido-bonded to tri(meta)phosphate. *Archs Sci. Genève* 51: 187-193.

VAN WAZER, J.R. 1958. Phosphorus and its Compounds, vol. I. Interscience, New York.

VOGEL, H.J. 1984. ³¹P-NMR studies on phosphoproteins. *In: Phosphorus-³¹P NMR Principles and Applications*. Pp. 105-153. Acad. Press, Inc, New York.

WESTHEIMER, F.H. 1987. Why nature chose phosphates. Science 235: 1173-1177.

YAMANAKA, J., K. INOMATA & Y. YAMAGATA. 1988. Condensation of oligoglycines with trimeta- and tetrametaphosphate in aqueous solutions. Orig. Life Evol. Biosph. 18: 165-178.