Characterization of "Trypanozoon" stocks by isoelectric focusing and isoenzyme analysis

Autor(en): Betschart, B. / Wyler, R. / Jenni, L.
Objekttyp: Article

Zeitschrift: Acta Tropica

Band (Jahr): 40 (1983)
Heft 1

PDF erstellt am: 26.10.2018

Persistenter Link: http://doi.org/10.5169/seals-313109

Nutzungsbedingungen
Die auf der Plattform e-periodica veröffentlichten Dokumente stehen für nicht-kommerzielle Zwecke in Lehre und Forschung sowie für die private Nutzung frei zur Verfügung. Einzelne Dateien oder Ausdrucke aus diesem Angebot können zusammen mit diesen Nutzungsbedingungen und den korrekten Herkunftsbezeichnungen weitergegeben werden.

Haftungsausschluss
Alle Angaben erfolgen ohne Gewähr für Vollständigkeit oder Richtigkeit. Es wird keine Haftung übernommen für Schäden durch die Verwendung von Informationen aus diesem Online-Angebot oder durch das Fehlen von Informationen. Dies gilt auch für Inhalte Dritter, die über dieses Angebot zugänglich sind.

Ein Dienst der ETH-Bibliothek
ETH Zürich, Rämistrasse 101, 8092 Zürich, Schweiz, www.library.ethz.ch

http://www.e-periodica.ch
Characterization of Trypanozoon stocks by isoelectric focusing and isoenzyme analysis

B. Betschart, R. Wyler, L. Jenni

Summary

Isoelectric focusing on ultrathin polyacrylamide foils, coupled with isoenzymatic analysis using five different enzymes, was used to characterize blood-forms of T. b. brucei, T. b. rhodesiense and T. b. gambiense and procyclic forms of T. b. brucei and T. b. rhodesiense. The protein concentrations in the lysates were quantified. Qualitative as well as quantitative differences were found.

Key words: isoelectric focusing; isoenzyme analysis; Trypanosoma b. brucei, T. b. rhodesiense, T. b. gambiense.

Introduction

Trypanosomes can be characterized by serotyping, DNA-analysis, and/or by their isoenzymatic patterns. Zymodemes have usually been determined by using starch gel- or cellulose-acetate-electrophoresis (Gibson et al., 1980; Kreutzer and Sousa, 1981): the advantage of cellulose acetate- over starch gel electrophoresis for T. cruzi has been shown by Lanham et al. (1981). Ebert (1982) introduced isoelectric focusing in thin layer polyacrylamide and agarose gels to characterize T. cruzi stocks. Lysates from twelve stocks were compared using seven different enzymes.

This paper extends the use of isoenzymatic analysis by isoelectric focusing to African Trypanozoon stocks.

Material and Methods

T. b. brucei (Clone STIB 247-F) derived from a primary isolate STIB 247, isolated from a hartebeest in the Serengeti National Park, Tanzania in 1971. The clone was selected from an ICR white mouse after 24 successive cyclical transmissions of STIB 247 through Glossina m. morsitans.

T. b. rhodesiense (Clone STIB 389-F) derived from a primary isolate STIB 389, isolated from man in Northern Tanzania in 1980. The clone was selected from an ICR white mouse after five successive cyclical transmissions of STIB 389 through G. m. morsitans.

Correspondence: Dr. B. Betschart, Swiss Tropical Institute, Socinstrasse 57, CH-4051 Basel
T. b. gambiense (STIB 387-A) was originally isolated by Dr. D. Mehlitz (Hamburg) from man at the Ivory Coast in 1978 as TH3/78E (031). The trypanosomes were harvested from Microtus montanus after one cyclical transmission through G. p. gambiensis.

Procyclic forms of T. b. brucei and T. b. rhodesiense were grown in medium SDM-79 (Brun and Schönenberger, 1979).

The bloodstream trypanosomes were separated from blood on a DEAE column (Lanham and Godfrey, 1970). The parasites were counted in a Neubauer chamber and finally pelleted at 1400 g for 10 min at 2-4°C. They were then lysed by adding an equal amount (v/v) of an aqueous solution of 2 mM dithiothreitol and 2 mM EDTA by three cycles of freezing on dry ice and thawing. The lysates were spun at 10,000 g for 30 min. The supernatant volumes were measured with a Hamilton syringe and the protein concentration estimated by a modified Lowry method (Peterson, 1977). The lysates were immediately frozen in aliquots as beads in liquid nitrogen.

Isoelectric focusing (IEF) was carried out using Servalyt Precotes pH 3-10 (Serva, Heidelberg). The foils were handled as described in the manual. They were placed on an LKB-Multiphor cooled to 8°C. The samples were loaded as 5 µl quantities, diluted to the appropriate protein concentrations (see results) into the applicator slits. Focusing was done with a PS 10 A Multistab (W. Meyer, Luzern) with a limited voltage of 1000 V. The current was restricted to 5 mA per foil and the focusing was stopped after 3½ hours. The foil was then cut into two pieces, and one was immediately overlayered with agarose and enzyme substrates and incubated at 37°C. The rest of the foil was used to determine the pH-gradient with a surface electrode (Ingold, Zürich).

The incubation mixtures for the different enzyme reactions were as described by Gibson et al. (1978). Five enzymes were used to characterize the trypanosomes: phosphoglucomutase (PGM) (E. C. 2.7.5.1), isocitrate dehydrogenase (ICD) (E. C. 1.1.1.42), aspartate aminotransferase (ASAT) (E. C. 2.6.1.1), alanine aminotransferase (ALAT) (E. C. 2.6.1.2) and aminopeptidase B (E. C. 3.4.11.1).

**Results**

To obtain a quantitative comparison between the lysates, the total number of trypanosomes was correlated with the protein concentration in each case. The protein concentrations had to be adjusted for the determination of each enzyme. Protein concentrations were adjusted to 1 mg/ml for ALAT, and to 5 mg/ml for ASAT. 10 mg/ml were needed for adequate staining of PGM, ICD and PEP. The lysates from procyclic forms had always to be used at half the above concentrations. These dilutions were necessary to avoid the overloading of the gel. The number of trypanosomes needed to get reproducible isoenzyme patterns ranged from 0.7-2.5×10⁷ trypanosomes, depending on the type of trypanosome populations as well as the enzyme.

We were able to discriminate between T. b. brucei and T. b. gambiense by the pattern obtained with PGM (Fig. 1a Nr. 1 and 3). T. b. gambiense shows a band focused at a pI of 5.7, whereas T. b. brucei shows two strong bands at pI 5.6 and 5.5. With ICD, T. b. gambiense (Fig. 1b Nr. 3) has a band at a pI 5.5, which is not present in T. b. brucei, which shows, besides minor bands, a relatively strong one at pI 5.7. T. b. rhodesiense showed the same pattern as T. b. brucei, but quantitative differences were found: in all assays performed so far T. b. rhodesiense showed less enzyme activity than T. b. brucei, even when the same amount of protein was applied (see Fig. 1). The patterns for ASAT and PEP were not strikingly different, but there may be quantitative differences between T. b. brucei and T. b. gambiense.
With all enzyme patterns tested the bloodform lysates can be distinguished from the procyclic forms of *T. b. brucei* and *T. b. rhodesiense*. With PGM a third strong band appears at pI 5.4 (Fig. 1a Nr. 4/5). With ICD-staining, four more strong anodic bands appear, in contrast to the single band at pI 5.7 from the bloodforms. *T. b. brucei* has only a weak band at pI 6.3 with ASAT (Fig. 1c Nr. 4). A band in addition to those found with the bloodforms is found with PEP at pI 4.9.
ALAT did not give good enough resolution for any differences between the lysates to be detected.

Discussion

Analysis of enzyme polymorphism requires a system which is sensitive and gives reproducible and easily-characterized results. Starch gel and cellulose-acetate electrophoresis are helpful techniques for the identification of trypanosomes (Gibson et al., 1980), but they have some inherent disadvantages, since no internal markers are included, which makes the comparison of different results impossible. Using isoelectric focusing the enzyme bands are characterized by their isoelectric point (pI) and the amount of protein applied can be accurately determined, so that the isoenzymes can be compared quantitatively.

Using the qualitative and quantitative differences we were able to discriminate between the Trypanozoon stocks studied, using only four enzymes. We are now investigating the general applicability of the patterns described using a variety of different African stocks.

Acknowledgments

This study was supported in part by a grant from the 14. Postgraduate Course in Experimental Medicine and Biology (R. W.) and by grant 3.681–0.80 from the Swiss National Science Foundation. We acknowledge the technical support of Dr. R. Brun, Ms. C. Kunz and Ms. S. Steiger.