Zeitschrift:	Acta Tropica				
Herausgeber:	Schweizerisches Tropeninstitut (Basel)				
Band:	41 (1984)				
Heft:	3				
Artikel:	Maintenance of "Glossina morsitans morsitans" on antiserum to procyclic trypanosomes reduces infection rates with homologous and heterologous "Trypanosoma congolense" stocks				
Autor:	Maudlin, I. / Turner, M.J. / Dukes, P.				
DOI:	https://doi.org/10.5169/seals-313299				

Nutzungsbedingungen

Die ETH-Bibliothek ist die Anbieterin der digitalisierten Zeitschriften auf E-Periodica. Sie besitzt keine Urheberrechte an den Zeitschriften und ist nicht verantwortlich für deren Inhalte. Die Rechte liegen in der Regel bei den Herausgebern beziehungsweise den externen Rechteinhabern. Das Veröffentlichen von Bildern in Print- und Online-Publikationen sowie auf Social Media-Kanälen oder Webseiten ist nur mit vorheriger Genehmigung der Rechteinhaber erlaubt. <u>Mehr erfahren</u>

Conditions d'utilisation

L'ETH Library est le fournisseur des revues numérisées. Elle ne détient aucun droit d'auteur sur les revues et n'est pas responsable de leur contenu. En règle générale, les droits sont détenus par les éditeurs ou les détenteurs de droits externes. La reproduction d'images dans des publications imprimées ou en ligne ainsi que sur des canaux de médias sociaux ou des sites web n'est autorisée qu'avec l'accord préalable des détenteurs des droits. <u>En savoir plus</u>

Terms of use

The ETH Library is the provider of the digitised journals. It does not own any copyrights to the journals and is not responsible for their content. The rights usually lie with the publishers or the external rights holders. Publishing images in print and online publications, as well as on social media channels or websites, is only permitted with the prior consent of the rights holders. <u>Find out more</u>

Download PDF: 05.08.2025

ETH-Bibliothek Zürich, E-Periodica, https://www.e-periodica.ch

¹ Tsetse Research Laboratory, ODA/University of Bristol, Bristol BS18 7DU, U.K.

² MRC Biochemical Parasitology Unit, The Molteno Institute, University of Cambridge, Cambridge CB2 3EE, U.K.

Maintenance of *Glossina morsitans morsitans* on antiserum to procyclic trypanosomes reduces infection rates with homologous and heterologous *Trypanosoma congolense* stocks

I. MAUDLIN¹, M. J. TURNER², P. DUKES¹, N. MILLER²

Summary

Three experimental groups of male *Glossina morsitans morsitans* were infected at their first feed with procyclic forms of different stocks of *Trypanosoma congolense* and subsequently maintained on a diet containing rabbit antiserum to one of these stocks. Control flies were similarly infected but were then maintained on normal rabbit serum. Dissection of the flies 19–21 days post infection showed a reduction in both immature and mature infection rates in all groups fed on antiserum by comparison with control flies. These results suggest that vaccination with a single procyclic *T. congolense* strain could reduce transmission of both homologous and heterologous *T. congolense* strains which might, in certain epidemiological circumstances, provide an alternative or additional control method for African trypanosomiases.

Key words: *Glossina morsitans; Trypanosoma congolense;* infection rates; African trypanosomiasis; vaccination; control.

Introduction

The salivarian trypanosomes have evolved a mechanism for eluding the immune system of their mammalian hosts which involves the production of a succession of variable antigenic types (VATs (Vickerman, 1978). The antigenic properties of the bloodstream and metacyclic forms of the trypanosome reside in its surface coat, which is made up of glycoprotein molecules, the composition of which differs for each VAT (Cross, 1975). The expression and control of the

Correspondence: Dr. I. Maudlin, Tsetse Research Laboratory, Department of Veterinary Medicine, Langford House, Bristol BS18 7DU, U.K.

genes involved in the production of these antigenic variants has recently been the subject of intense research which has shown such variation results from genetic rearrangements within the trypanosome genome (Borst and Cross, 1982; Englund et al., 1982; Turner, 1982). It appears that this antigenic heterogeneity is acquired as soon as the metacyclic trypanosomes mature in the tsetse fly (LeRay et al., 1978). These findings have profound implications for the development of a vaccine designed to protect mammals against trypanosome infection, suggesting as they do that a very complex "cocktail" of antigens would be necessary to provide effective protection. The present work examines an alternative approach to trypanosomiasis control which would be to use a vaccine which elicits an antiserum effective against the procyclic trypanosomes found in the tsetse fly midgut. These trypanosomes have lost the antigenically variable protective surface coat (Barry and Vickerman, 1979) and become vulnerable to such an antiserum when ingested by the fly as part of a blood meal (Murray et al., 1980). Such an "altruistic" approach to vaccination, through transmission blocking immunity, has also been suggested for the control of malaria as anti-gamete antibodies raised in the mammalian host prevent gamete fertilization in the midgut of mosquitoes (Gwadz, 1976; Rener et al., 1980).

Materials and Methods

A single stock of Trypanosoma congolense (LUMP 82, derived from TREU 621 isolated in Uganda in the 1960's) in procyclic culture (Evans, 1979) was used to raise antiserum in a rabbit. The rabbit was immunized by intradermal injection with 0.5 ml of a 1% suspension of frozen $(-80 \degree \text{C})$ and thawed T. congolense procyclic forms in RPMI 1640 emulsified with an equal volume of Freunds complete adjuvant. Booster injections consisting of 0.5 ml of a 6% suspension of frozen and thawed T. congolense procyclic forms in RPMI 1640 emulsified with an equal volume of Freunds incomplete adjuvant were given 5 and 15 weeks after primary immunization. A test serum sample, taken at 17 weeks, was immunofluorescence test (Miller and Turner, 1980) positive on suspensions of T. congolense procyclic forms but not on similar suspensions of T. brucei. The rabbit was exsanguinated 24 weeks after primary immunization and the serum heat inactivated and freeze dried. After reconstitution in distilled water, the serum was sterilized by filtration and then diluted 1:1 with normal rabbit serum which had been similarly treated; control serum from untreated rabbits was heat inactivated and treated identically to the experimental serum. To determine whether this antiserum to T. congolense LUMP 82 procyclic forms could eliminate trypanosome infections in the vector, it was tested in flies against the homologous stock (LUMP 82) and two heterologous stocks of the same species: TRPZ 133 isolated from a dog in Zambia in 1981 and 1/148 FLY isolated from a cow in Nigeria in 1960.

Male *Glossina morsitans morsitans* in groups of 200 were infected, 18–30 h following emergence from the puparium, with procyclic culture forms of *T. congolense* in whole pig blood. In the first experiment (LUMP 82) the infecting dose was 6.4×10^6 trypanosomes/ml blood; in experiment 2 the dose of TRPZ 133 was 1.6×10^6 /ml and in experiment 3 the dose of 1/148 FLY was 6.4×10^6 /ml. Flies which refused the infective feed were removed from the experiment. For subsequent feeds flies were fed six days a week on a diet of one volume of pig red cells (washed $3 \times$ in phosphate buffered saline) mixed with two volumes of either rabbit antiserum to LUMP 82 (test flies) or normal rabbit serum (control flies). Flies were infected and maintained on an artificial membrane feeding system (Mews et al., 1977). 19–21 days post infection flies were dissected and the

gut, labrum and hypopharynx of each were examined for trypanosomes by phase contrast microscopy ($\times 400$).

Results

The results of the dissections are given in Table 1 which shows that antiserum raised against a single strain of T. congolense can significantly reduce infection rates in flies infected with heterologous T. congolense stocks as well as in those infected with the homologous stock. In experiment 1, using the homologous stock, gut and labral infection rates were significantly reduced in flies fed on antiserum when compared with the control flies. In addition, the mature (hypopharyngeal) infection rate was halved, although statistical significance was not demonstrable with the small numbers involved. [In a preliminary experiment with LUMP 82, in which control flies were fed foetal calf serum instead of rabbit serum, results were closely similar to those shown here; when pooled these data demonstrated a significantly reduced mature infection rate, from 14% (162 flies) in the controls to 5% (171 flies) in the experimental flies (γ^2 = 5.4, 0.05 > P > 0.01).] The second experiment produced only a small reduction in gut TRPZ 133 infection rates but a significant reduction in mature infections, from 21% in the control flies to 8% in the experimental flies. The results of the third experiment were similar to those of the first, with 1/148 FLY gut and labral infection rates significantly reduced compared with control flies; the reduction in maturation rate was not statistically significant, again probably due to the small numbers involved.

Table 1. Percentage infection rates in midgut (G), labrum (L) and hypopharynx (H) of three experimental groups of male *G. m. morsitans* infected with three different stocks of *T. congolense* and fed on diet containing rabbit antiserum to a single stock (LUMP 82) or, control diet containing normal rabbit serum. Gut (G) infection rates refer to the proportions of flies with trypanosomes in the midgut whether or not the mouthparts were also infected. Numbers of flies dissected in each group given in parentheses. Statistical comparisons by χ^2 tests

Expt. No.	Trypanosome		Diets		Probabilities
	stock		Experimental	Control	
1	LUMP 82	G G, L G, L, H	34 (89) 19 8	64 (92) 46 15	<0.001 <0.001 not significant
2	TRPZ 133	G G, L G, L, H	43 (95) 35 8	52 (95) 42 21	not significant not significant 0.01>P>0.001
3	1/148 FLY	G G, L G, L, H	22 (94) 2 0	52 (92) 13 3	<0.001 0.01>P>0.001

Discussion

The present work has shown that T. congolense infection rates can be significantly reduced by feeding flies on antiserum raised against a single stock of cultured procyclic trypanosomes. This is consistent with theoretical considerations which suggest that the procyclic culture form is equivalent to the gut form of the trypanosome, having an antigenically invariable surface, unlike bloodstream and metacyclic trypomastigotes (Vickerman, 1970). Murray et al. (1982) have recently shown that antibody raised against a single T. congolense procyclic stock (ILRAD C24) will reduce infection rates in flies infected with the same and different stocks of T. congolense. The present results show that antiserum to a single procyclic stock affects both the establishment of procyclic infections and the subsequent maturation of both homologous and heterologous stocks in tsetse. Reduced procyclic infection rates may result directly from antibody lysis of trypanosomes, or indirectly from the action of tsetse gut enzymes or serum lipids on antibody-agglutinated trypanosomes (Maudlin et al., 1984). It is likely that reduced maturation rates follow from inactivation of trypanosomes moving forward to the proboscis; alternatively, antibody may inhibit trypanosome morphogenesis as described in Trypanosoma cruzi (Sher and Snary, 1982).

These results also point to a possible alternative method of trypanosomiasis control, in particular of sleeping sickness if similar results can be obtained with *T. brucei gambiense*. For example, in parts of the Ivory Coast where *G. palpalis palpalis* may take more than 80% of their feeds from domestic pigs (Gouteux et al., 1982), trypanosome infection rates in the fly population might be reduced significantly by vaccinating domestic livestock with a simple procyclic antigen. Similarly, for bovine trypanosomiasis, the trypanosome "challenge" (Whiteside, 1958) in a ranching area might be significantly reduced by vaccinating herds of cattle with a procyclic antigen and we are examining this by computer simulation. The number and frequency of feeds that infected flies must take from animals immunized with procyclic antigen to reduce infection rates significantly, and the frequency of host vaccination required to maintain effective antiserum levels, are currently being investigated; clearly these are likely to be important determinants of the efficacy of a procyclic vaccine.

Acknowledgments

We should like to thank D. A. Evans and C. M. Scott of the London School of Hygiene and Tropical Medicine for trypanosome stocks and Malcolm Flood for technical assistance. The financial support of the Overseas Development Administration of the Foreign and Commonwealth Office to the Tsetse Research Laboratory is gratefully acknowledged.

Borst P., Cross G. A. M.: Molecular basis for trypanosome antigenic variation. Cell 29, 291–303 (1982).

Barry J. D., Vickerman K.: *Trypanosoma brucei:* loss of variable antigens during transformation from bloodstream to procyclic forms in vitro. Exp. Parasit. 48, 313–324 (1979).

- Cross G. A. M.: Identification, purification and properties of clone-specific glycoprotein antigens constituting the surface coat of *Trypanosoma brucei*. Parasitology *71*, 393–417 (1975).
- Englund P. T., Hajduk S. L., Marini J. C.: The molecular biology of trypanosomes. Ann. Rev. Biochem. 51, 695–726 (1982).
- Evans D. A.: Cyclical transmission of *Trypanosoma brucei rhodesiense* and *Trypanosoma congolense* by tsetse flies infected with culture-form procyclic trypanosomes. J. Protozool. *26*, 425–427 (1979).
- Gouteux J. P., Laveissière C., Boreham F. L.: Ecologie des glossines en secteur pré-forestier de Côte d'Ivoire. 3. Les préférences trophiques de *Glossina pallicera* et *G. nigrofusca*. Comparaison avec *G. palpalis* et implications épidémiologiques. Cah. O.R.S.T.O.M. sér. Ent. méd. Parasit. 20, 109–124 (1982).
- Gwadz R. W.: Malaria: successful immunization against the sexual stages of *Plasmodium gallinaceum*. Science 193, 1150–1151 (1976).
- LeRay D., Barry J. D., Vickerman K.: Antigenic heterogeneity of metacyclic forms of *Trypanosoma* brucei. Nature (Lond.) 273, 300–302 (1978).
- Maudlin I., Kabayo J. P., Flood M. E. T., Evans D. A.: Serum factors and the maturation of *Trypa-nosoma congolense* infections in *Glossina morsitans*. Z. Parasitenk. 70, 11–19 (1984).
- Mews A. R., Langley P. A., Pimley R. W., Flood M. E. T.: Large-scale rearing of tsetse flies (*Glossina* spp.) in the absence of a living host. Bull. ent. Res. 67, 119–128 (1977).
- Miller E. N., Turner M. J.: Analysis of antigenic types appearing in first relapse populations of clones of *Trypanosoma brucei*. Parasitology 82, 63–80 (1980).
- Murray M., Barry J. D., Morrison W. I., Williams R. O., Hirumi H., Rovis L.: A review of the prospects for vaccination in African trypanosomiasis. Wld Anim. Rev. 33, 14–18 (1980).
- Murray M., Moloo S. K., Hirumi K., Dar F., Hinson C. A., Hirumi H.: Reduction of trypanosome infection in tsetse flies by antibodies raised against procyclic forms of *Trypanosoma congolense*. Proc. Fifth Int. Congress Parasit. Toronto 2, 103 (1982).
- Rener J., Carter R., Rosenberg Y., Miller L. H.: Anti-gamete monoclonal antibodies synergistically block transmission of malaria by preventing fertilization in the mosquito. Proc. nat. Acad. Sci. (Wash.) 77, 6797–6799 (1980).
- Sher A., Snary D.: Specific inhibition of the morphogenesis of *Trypanosoma cruzi* by a monoclonal antibody. Nature (Lond.) *300*, 639–640 (1982).
- Turner M. J.: Biochemistry of the variant surface glycoproteins of salivarian trypanosomes. Advanc. Parasit. 21, 69–153 (1982).
- Vickerman K.: Morphological and physiological considerations of extracellular blood protozoa. In: Ecology and physiology of parasites, ed. by A. M. Fallis, p. 58–91. Adam Hilger, London 1970.
- Vickerman K.: Antigenic variation in trypanosomes. Nature (Lond.) 273, 613-617 (1978).
- Whiteside E. F.: The maintenance of cattle in tsetse-infested country. A summary of four years' experience in Kenya. Int. Sci. Committee Trypano. Res. 7th Meeting, Brussels, p. 83–90 (1958).